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Effects of Liver Tissue Transplant on Finger Regeneration in Mouse

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Abstract:

Finger regeneration ability in mammals is very poor, mammals which were designated as non-regenerating models among vertebrates also can be made to regenerate the lost structures if conditions favoring regeneration are provided, near the amputated site particularly during the early post amputational periods.

It can be induced by Allografting of liver tissue at amputated finger. Studies carried out on transplantation of liver tissue near the wound surface after amputation of finger through middle phalanx have shown many interesting results.

Such findings would definitely show the way to bio-medical scientists to search means, so that human beings suffering from amputational injuries, with permanent disabilities can be provided reparative therapy with the help of tools of tissue engineering.

Keywords: Regeneration, amputation, liver tissue, transplantation

1. Introduction

Restitution of the missing structure by the rest of an organ or body part of an animal is known as regeneration. (Wallace-1981).

Mammals were thought to possess very poor or no regenerative abilities. Illingsworth in 1974, reported about regeneration of lost or crushed fingertips in young children in certain clinical cases, provided amputation level was in front of the distal most inter-phalangeal joint. In these cases, restitution of fingertips was observed within 3 to 4 months provided treatment of wound was restricted to aseptic dressings only. Whenever, the wound was sutured with the skin over the wound surface, it generally resulted in failure of regeneration. There was no regeneration when the level of amputation was more proximal i.e. through the mid phalanx.

In higher animals including human beings many wear and tear processes continue from beginning of body development until death. The outer layer of skin, Blood cells, Sperm cells, uterine lining, Intestinal mucosa etc. are replenished as per requirement. Another reparative phenomenon that occurs in higher mammals is the acquisition of functions by existing tissues, when part of the organ ceases to function in certain disease conditions or is removed.

Sharma, 2002 reported that traumatization of amputated finger, improved quality of regeneration by improving bone growth and formation of healthy cartilage at distal end Extensive studies on regenerative ability of mouse finger were carried out in the laboratory of Sharma (1988 & 1991) at the MDS University Ajmer (India) who reported regeneration of distal part of spatula in mouse at embryonic stages using intra amniotic surgical techniques and embryo culture techniques and found that finger amputation through distal part of the autopodium at 16th day of gestation stage of embryo show complete regeneration (Archana, 1992). Amputation through the distal phalanx in adult mice resulted in complete restitution of lost limb elements confirming the results of Borgens (1982). It was found that those cases where hyaline cartilage of head of middle phalanx or base of distal phalanx persists normally show good regeneration.

Small-for-size liver allografts without immune suppression have decreased survival compared with full-for-size grafts for the concomitant regeneration-induced accelerated rejection. This study was designed to examine the effect of zinc finger protein A20 on liver allograft regeneration and acute rejection using a high responder rat model of 30% partial liver transplantation (Xu et al., 2009).

Yang et al.,(2012) reported that allogeneic liver transplantation from DA rats to Lewis rats was performed. Chronic liver allograft dysfunction was induced in the rats by administering low-dose tacrolimus at postoperative day 5.

It has been mentioned that mouse show a very limited ability to regenerate finger elements. This too is restricted to regeneration of fingertips after amputation injury at the distal most phallanx. In the recent years Molecular Biology researches in this area have indicated that regeneration can be induced in non-regenerative cases if suitable environment is provided to the cells, which help in regeneration. The environmental tissues of the stump allow the formation of cartilage and bone, and this is proposed that limb formation could be possible by supplying competent cells endogenously or exogenously.

Fernando et al., (2011) studied that amputation of the distal region of the terminal phalanx of mice causes an initial wound healing response followed by blastema formation and the regeneration of the digit tip. Thus far, most regeneration studies have focused in embryonic or neonatal models and few studies have examined adult digit regeneration.

We are faced with a limb-severing injury, any regenerative attempt would endeavour to accelerate the pace at which the tissue heals to a clinically relevant/functional state. The science of limb regeneration can be approached from three different angles, developmental biology; regenerative medicine; and tissue engineering (See et al., 2013).

A more contemporary approach to the stimulation of regeneration is the application of cellular engineering principles, which involve strategies such as the implantation of the cultured cells, with or without appropriate substrate. Among the many tissues in mammals, bone has considerable power of regeneration and is a prototype model for tissue engineering based on morphogenesis. As concluded by the studies carried out by the scientists of center for tissue regeneration and repair, University of California, Implantation of demineralized bone matrix into subcutaneous sites results in local bone induction. Hand/upper extremity transplantation is the most common form of vascularized composite allotransplantation performed to date. An Update of worldwide outcomes is reported (Shores et al., 2015).

In mammals, liver possess very high regenerative abilities. So, we transplanted liver tissue taken from the liver of an adult mouse to the amputated finger in another group of animals. The results obtained from the above studies clearly demonstrate that liver tissue can also induce regeneration in the non-regenerating cases.

In view of this the objective is to find means to induce regeneration in otherwise non-regenerating cases by liver tissue transplantation.

2. Materials and Methods

Experiments related to proposed study were carried out on closely inbred *Swiss albino* mice. Studies consisted of the following main lines of research.

1. To study the cellular and tissue organization of the finger in the distal and middle phalanx to find the precise site for transplantation of inducing factors.
2. To find means to induce regeneration in otherwise non-regenerating cases because normally amputation through middle phalanx in adult mouse does not show any regeneration.
3. Attempt to induce regeneration, by transplantation of certain tissue with known developmental potencies. e.g. liver tissue.

The amputated animals were kept separated from the breeding colony in the animal house and germ free and aseptic conditions were maintained.

The 3rd finger of hind limb of 4-week-old mouse is about 5 mm long including nail. Skeleton of finger is well, ossified. Dense hair is present on the dorsal side of finger. The sagittal and longitudinal section (Fig. 1 & 2) of finger shows three phalanxes namely distal phalanx, middle phalanx and proximal phalanx. Nail is present at the distal phalanx and is curved.

2.1. Amputation Procedure

Adult mice (4 week \pm 1 days¹) were anesthetized by placing them in a sterile chamber where diethyl ether soaked cotton was placed near the nostrils of the experimental animal. It took about ½ to 2 minutes for anesthetization. Third finger of hind limbs were amputated using a sterile surgical blade under surgical microscope below the head of the middle phalanx.

2.2. Experimental Design

Transplantation experiments were carried out on 4-week-old Swiss albino mice. Following amputation through the shaft of the middle phalanx, animals were divided into the following groups

2.2.1. Group I--Control Group

After amputation animals of control group were left for healing under clean and sterile conditions in cages without any further treatment. Observations were made on 60th day post amputation

2.2.2. Group II—Tissue Transplanted treated group

The tissue was transplanted in animals seven days after amputation of middle finger. The piece of liver tissue of the 4-week-old mouse and was transplanted

2.3. Transplantation Procedure

The transplantation was performed under aseptic conditions. First of all, an incision was made on the latero-ventral side of finger just near the base of the middle phalanx. A tunnel was made through it and the tissue to be transplanted was pushed to the amputated shaft of the middle phalanx

The care was taken that the transplant does not come out of the tunnel. The transplantations were made after 7th day of amputation. The post amputational developments after transplantation were examined in the control and transplanted groups on the 60th day for morphological developments.

3. Morphological and Histological Analysis

For the analysis of control and treated post amputational development (morphology and histology) fingers (Fig. 3 & 4) amputated through the shaft of middle phalanx were fixed in Bouin's solution.

3.1. Observation and Results

3.1.1. Control

On the 5th day after amputation, healing was in progress wound epidermis arise from the cut margins towards the center and healed up the wound surface. Scab material was present at the apical end of the stump. On the 10th day post amputation, the stump showed disappearance of scab. The wound surface was covered by the wound epidermis. On the 40th day the amputated finger stump shows simple healing of wound epidermis. The wound healing was completed by the 60th day post amputation. The stump surface was flattened and was covered with dense hair (Fig. 5 & 6). The distal most ventral ridge was not regenerated and the thick dermal pad was also not visible as supported by histological studies (Fig. 7)

3.2.2. Treated

Liver tissue from the 4-week-old mouse was transplanted in this set of organisms on 10th day post transplantation. Complete wound healing with partial regeneration of fingertip had taken place. The wound skin was not yet covered with the hair. By the 40th day after transplantation the regeneration of the ventral ridge is completed and by the 60th day growth of nail has also started. Histological observation on the 60th day revealed repair and restitution of component of fingertip at the cut end of the stump. Skin of the stump was well formed and epidermal derivatives were also visible in adjacent dermal region. The sub-epidermal region was filled with fibroblast cells. Internal organization and nail structure showed complete restitution of amputated part of the finger.

3.2. Conclusion

Studies carried out on transplantation of liver tissues near the wound surface after amputation of finger through middle phalanx have shown many interesting results. The stumps, which do not normally regenerate after amputation through middle phalanx, show restoration of normal pattern in many cases with allograft of liver tissue taken adult mouse.

3.3. Acknowledge

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4. References

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Appendix

Morphological and histological figures of middle finger of hind limb of 4 week old mouse

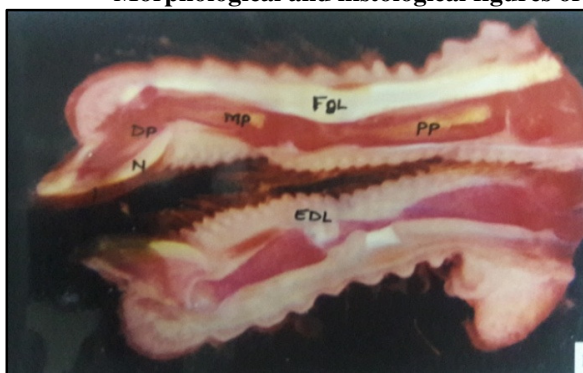


Figure 1: Saggital Section of middle finger (20X)

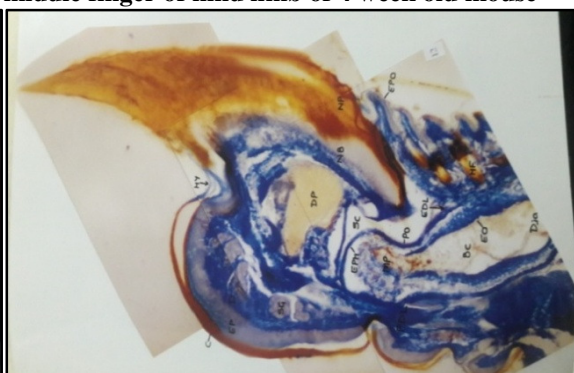


Figure 2: Longitudinal section of middle finger (40X)

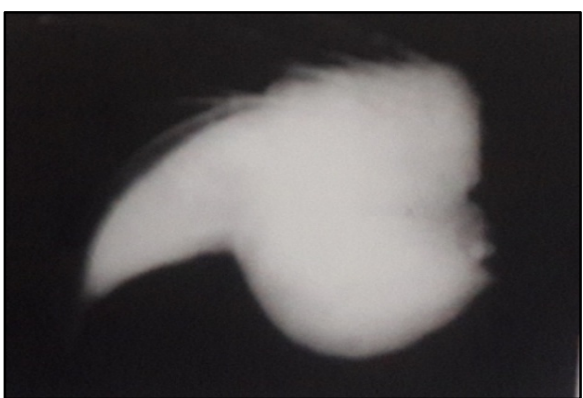


Figure 3: Morphology of removed part of finger tip (15X)

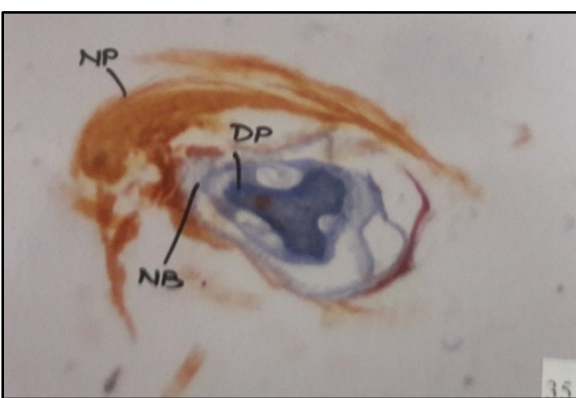


Figure 4: Cross section of finger tip passing through DP (10X)



Figure 5: Post-amputated finger after 60 days (15X)



Figure 6: Post-amputated finger removed after 60 days (15X)

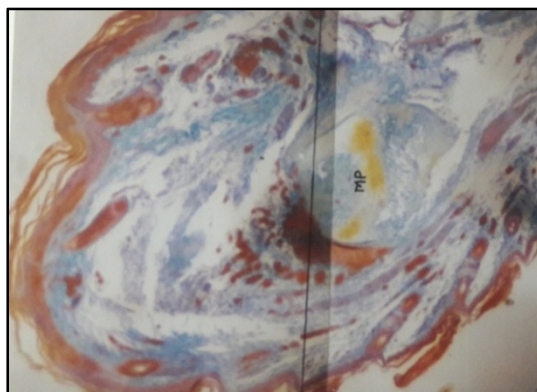


Figure 7: L.S. of post-amputated finger removed after 60 days (40X)



Figure 8: Morphology of amputated finger 60 days Post transplantation (Lateral View 15 X)

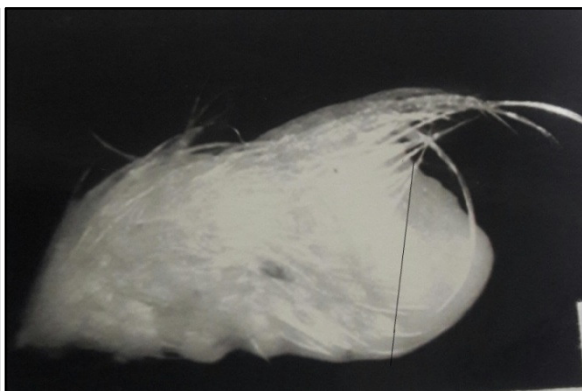


Figure 9: Morphology of amputated finger removed after 60 days Post transplantation (15X)



Figure 10: LS showing regeneration of head of middle phalanx and distal phalanx as observed on 40 day

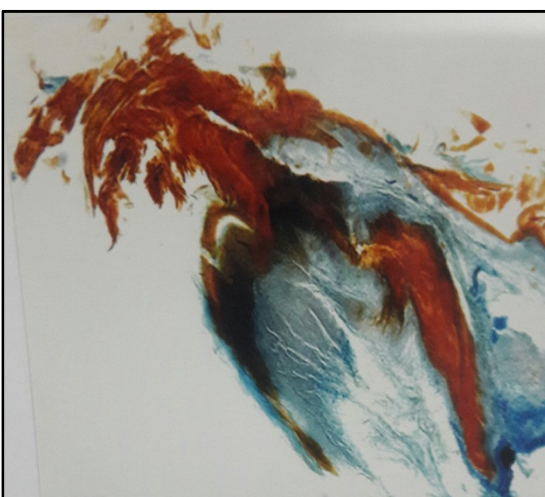


Figure 11: LS showing regeneration of skeletal elements and nail as observed on 60th day post transplantation (40X)

BC- Bone cavity
 DP – Dispal phalanx
 EO- Endosteum
 HY- Hyponychium
 PO- Periosteum
 FDL – Flexor digitorum longus tendon

D- Dermis
 EPH- Epiphysis
 EP – Epidermis
 NB- Nail bed
 SC- Synovial Cavity

Dia – Diaphysis
 EPH- Epiphysis
 HF- Hair follicle
 NP- Nail Plate
 SG-Sweat Gla
 EDL- Extesor Digitorum longus tendon