



A Comparative Study And Evaluation Of The Diagnostic Utility Of Adenosine Deaminase (ADA) Activity In Pleural, Peritoneal And Cerebrospinal Fluid For The Diagnosis Of Tuberculosis

Dr. Madhulika Mistry

Assistant Professor, MD, Department of Microbiology, P.D.U. Medical College, Rajkot, India

Yogesh Goswami

Professor, Department of Microbiology, P.D.U. Medical College, Rajkot, Gujarat, India

Mehta Krunal

Tutor, MD, Microbiology, Department of Microbiology, P.D.U. Medical College, Rajkot, India

Krupali Gadhvi

1st year Resident, Department of Microbiology, P.D.U. Medical College, Rajkot, India

Abstract:

Background Tuberculosis is a major cause of effusion & meningitis. Diagnosis by conventional methods has either less sensitivity or time consuming. Detection of ADA enzyme activity has been proposed to be a useful surrogate marker for diagnosis of tuberculous effusion & TB meningitis.

Objective To Assess the value of ADA activity in aspirated pleural, peritoneal and cerebrospinal fluid for the diagnosis of TB infection.

Methods Total 150 fluid; 70 pleural, 16 peritoneal and 64 cerebrospinal fluid were evaluated for ADA activity along with routine biochemical, bacteriological and cytological examination. Findings are correlated with clinical and radiological evidences.

Results Among confirmed 51 TB pleural effusion: ADA activity found positive (> 60 U/L) in 46 (90.2%) borderline (between 40 to 60 U/L) in 4. only 1 (1.96%) was negative (< 40 U/L).

Among 2 TB peritoneal effusions: ADA activity found positive in both (100%)

Among confirmed 21 TB meningitis cases: ADA activity found positive (> 10 U/L) in 20 (95.2%) and only 1 (4.8%) was negative (< 10 U/L).

Conclusion Diagnosis of tuberculous effusion by direct smear, culture, cytology and biochemical tests have either limited sensitivity or specificity while test for biochemical marker- ADA activity is simple, rapid, and economic and has significant role in early diagnosis of tuberculous effusion and TB meningitis.

Introduction

Despite the discovery of tubercle bacillus more than a hundred years ago, and all the advances in our knowledge of the disease made since then, tuberculosis still remains one of the major health problems facing mankind, particularly in developing countries. Presently about one third of the world's population is infected with mycobacterium tuberculosis. It is estimated that currently there are about 10 million new cases of tuberculosis every year with 3 million deaths occurring world-wide⁽¹⁾

Currently more people die of tuberculosis than from any other infectious diseases. nearly 95% of all tuberculosis cases and 98% of deaths due to tuberculosis are in developing countries and 75% of tuberculosis cases are in the economically productive age group⁽²⁾. In India, out of a total population over 1 billion, each year about 2 million develop active disease and up to half million die⁽³⁾. It also imposes a cost on our economy in terms of current and future output losses because of premature deaths & ill health⁽⁴⁾.

To add to the existing burden, the situation is compounded by large scale increase of new TB cases associated with increasing HIV-infection. India is estimated to have 3.5 million HIV patients and about 1.8 million of these are co-infected with TB⁽⁵⁾.

Early diagnosis of tuberculosis and initiating optimal treatment would not only enable a cure of individual patient but will also curb the transmission of infection and diseases to others in community. Diagnosis of tuberculosis confirmed mainly by direct sputum examination of AFB and by culture. But diagnosis of tuberculosis pleural effusion and TB meningitis are difficult due to low sensitivity of the various standard diagnosis tools. The diagnosis can be established by demonstrating elevated level of Adenosine Deaminase (ADA) or interferon gamma in the aspirated fluid⁽⁶⁾.

However the diagnoses of tuberculous effusion require investigation of fluid biochemistry, cytology and pleural biopsy. Positivity for AFB is very low and culture is time consuming. ELISA, PCR and interferon are very expensive test.

Adenosine Deaminase (ADA) has been proposed to be a useful surrogate marker for the diagnosis of tuberculosis in pleural, pericardial and peritoneal fluids. Studies have confirmed high sensitivity and specificity of Adenosine Deaminase for early diagnosis of extra pulmonary tuberculosis^(7, 8, 9).

ADA is an enzyme widely distributed in mammalian tissue particularly in T lymphocytes. ADA is enzymes that catalyses conversion of Adenosine to inosine. Adenosine is a predominant T lymphocytes enzyme and its plasma activity is high in

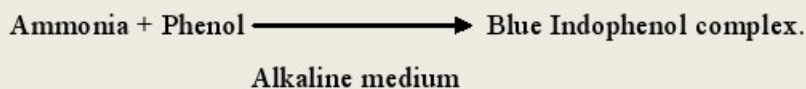
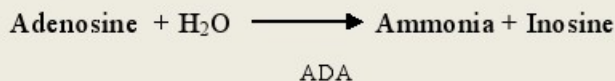
diseases in which cellular immunity is stimulated. Increase level of ADA found in various forms of tuberculosis making it a marker for the same ⁽¹⁰⁾.

Materials And Methods

This study was carried out on 150 patients suffering from pleural, peritoneal effusion and meningitis who attended OPD or were admitted in P.D.U. Government Medical College & Hospital, Rajkot during April to July 2012. Detailed clinical history, physical examination and investigations e.g. smear for AFB, cytological examination, biochemical examination and wherever possible USG, X-ray chest and other appropriate investigations including ADA level in aspirated fluid were carried out in all the patients. ADA estimation was done by commercially available kits based on colorimetric method with following principles.

Principle

Adenosine Deaminase hydrolyses adenosine to ammonia and inosine. The ammonia formed, further reacts with phenol and hypochlorite in an alkaline medium to form a blue indophenols complex with sodium nitropruside acting as a catalyst. Intensity of blue colored complex formed is directly proportional to the amount of ADA present in sample.



Reference value

| Specimen | Reference value (ADA) |
|--|---|
| Serum, plasma, Pleural, Pericardial Peritoneal fluid | Normal < 40 U/L, Positive > 60 U/L Suspect > 40 U/L to < 60 U/L, |
| CSF | Normal < 10 U/L, Positive > 10 U/L |

Table 1: Reference value (ADA)

Test Procedure

| | Blank(ml) | Standard(ml) | Sample blank(ml) | Test(m) |
|--------------------------|-----------|--------------|------------------|---------|
| Buffer | 0.20 | 0.20 | - | - |
| Reagent | - | - | 0.20 | 0.20 |
| Adenosine reagent | 0.02 | - | - | - |
| Deionised water | | | | |
| Standard | - | 0.02 | - | - |
| Sample | - | - | - | 0.02 |

Table 2

Mix well incubated 37°C for 60 minutes and then adds the following:

| | | | | |
|------------------------------|-----|-----|------|-----|
| Working phenol reagent | 1.0 | 1.0 | 1.0 | 1.0 |
| Sample | - | - | 0.02 | - |
| Working Hypochlorite reagent | 1.0 | 1.0 | 1.0 | 1.0 |

Table 3

Mix well incubated 37°C for 15 minutes or at R.T. for 30 minutes.

Measure the absorbance of blank, standard, sample blank and test against distilled water.

Calculations:

$$\text{Total ADA activity In U/L} = \frac{\text{Absorbance of Test} - \text{Absorbance of SB}}{\text{Absorbance of S} - \text{Absorbance of B}} \times 50$$

Linearity

The Procedure is Linear up to 150 U/L if the values exceeded this limit dilute the sample with deionised water and repeat this assay. Calculate the value using appropriate dilution factor.

Result And Observations

| Type of fluid | NO. | ADA activity in U/L | | |
|----------------------------|-----|---------------------|------------------------------|----------------------|
| | | normal < 40 U/L | suspect >40 U/L to <60U/L | positive >60 U/L |
| Pleural fluid | 70 | 17 | 06 | 47 |
| Ascitic (Peritoneal fluid) | 16 | 13 | 01 | 02 |
| CSF | 64 | Normal<10 U/L 44 | – | Positive>10U/L 20 |
| Total | 150 | 74 | 07 | 69 |

Table 4

Out of total 150 fluid specimens; 70 were pleural fluid, 16 were peritoneal fluid and 64 were cerebrospinal fluid. Of 70 pleural fluid tested for ADA activity; 48 showed ADA activity > 60 U/L (positive), 14 showed activity < 40 U/L (normal) and 08 showed ADA activity between 40 -60 U/L (suspect). Of 16 peritoneal fluid tested; 2 showed positive, 13 showed negative & 1 showed suspect result. Of 64 CSF tested; 20 showed ADA activity > 10 U/L (positive) and 44 showed activity < 10 U/L (normal).

| Type of fluid | Diagnosis | NO | ADA activity | | |
|---------------------|--------------|----|--------------|------------|------------|
| | | | Positive | Normal | Suspect |
| Pleural Fluid 70 | Tuberculosis | 51 | 46 (90.2%) | 04(7.8%) | 01(2%) |
| | non TB | 19 | 01 (5.3%) | 02 (10.5%) | 16 (84.2%) |
| Peritoneal fluid 16 | Tuberculosis | 02 | 02 (100%) | 00 | 00 |
| | Non TB | 14 | 00 | 01 (7.2%) | 13 (92.8%) |

Table 5

| Type of fluid | Diagnosis | No. | ADA activity | |
|---------------|--------------|-----|--------------|-----------|
| | | | Positive | Negative |
| | | | >10 U/L | <10U/L |
| CSF 64 | Tuberculosis | 21 | 20 (95.2%) | 01 (4.8%) |
| | Non TB | 43 | 00 | 43 (100%) |

Table 6

On correlating the clinical, radiological, cytological & biochemical findings, out of total 70 pleural effusion patients; 51 were diagnosed as tuberculous effusion of which 46 (90.2%) showed increased ADA activity, 1 (2%) showed normal ADA activity & 4 (7.8%) showed suspect results. From 19 non TB pleural effusions 16 (84.2%) showed normal activity. Of total 16 peritoneal effusion patients; two diagnosed as tuberculosis origin and both (100%) showed increased ADA activity. Of 14 non tuberculosis patients, 13 (92.8%) showed normal ADA activity (<40 U/L). Out of total 64 CSF specimen tested; 21 diagnosed as tuberculous meningitis of which 20 (95.2%) showed increased ADA activity. All 43 patients having non tuberculous etiology showed normal ADA activity (100%) in CSF samples.

So overall 74 patients diagnosed having tuberculosis; 68 (92%) showed increased ADA activity, 2 (2.7%) showed normal ADA activity and 4 showed suspect results. If we take 50 U/L as cut off point, 72 (97.3%) showed increase ADA activity.

| Specimen | Diagnosis as TB | AFB positive |
|------------------|-----------------|--------------|
| Pleural fluid | 51 | 03 (5.9%) |
| Peritoneal fluid | 02 | 00 (0%) |
| CSF | 21 | 01(4.8%) |
| Total | 74 | 04 (5.4%) |

Table 7

AFB positivity by ZNCF technique as per the RNTCP guideline in specimens suggests very low sensitivity of direct smear examination for AFB in diagnosis of tuberculous effusion.

| Age | No. of patients having TB etiology | | | Total |
|--------------|------------------------------------|------------------|-----------|-----------|
| | Pleural fluid | Peritoneal fluid | CSF | |
| 00-10 Years | 08 | 00 | 03 | 11 |
| 11-20 Years | 05 | 00 | 03 | 08 |
| 21-30 Years | 14 | 01 | 03 | 18 |
| 31-40 Years | 13 | 01 | 02 | 16 |
| 41-50 Years | 04 | 00 | 02 | 06 |
| 51-60 Years | 02 | 00 | 02 | 04 |
| >60 years | 05 | 00 | 00 | 05 |
| Total | 51 | 02 | 15 | 68 |

Table 8

In this study it is found that tuberculous effusion predominantly occurs in adult patients: 20-40 years age group and tuberculous meningitis predominantly occur in pediatric and young patients.

ADA, a product of T lymphocytes has been reviewed as an excellent marker for the diagnosis of tuberculous pleural effusion. The raised ADA activity under antigenic stimulation is found in infections such as tuberculosis where cell mediated immunity (CMI) is stimulated⁽¹¹⁾.

Almost all research workers have shown sensitivity & specificity of 90% to 100% for the value ADA in pleural fluids using different cuts off levels.

Gupta D.K.⁽¹²⁾ studied 53 cases of pleural effusion out of which 36 were of TB etiology. The mean ADA level I tuberculosis was 50.75 U/L while in malignant and Para pneumonic effusion it was 14.47 U/L and 28.65 U/L respectively. The sensitivity & specificity for diagnosis tuberculosis were 100% & 94% respectively.

Burgess L.J. ⁽¹³⁾ showed ADA activity in tuberculous effusion was higher than in any other diagnostic group. At a level of 50 U/L the sensitivity and specificity for the identification of tuberculosis was 90% and 89% respectively.

Value of activity in pleural effusion was studied by Shibagaki T et al ⁽¹⁴⁾. He concluded that tuberculous pleural effusion had a much higher ADA activity than cancer effusion and total ADA activity in tuberculous pleural effusion decreases after anti tubercular therapy.

Using cut off level of 25 U/L in serum and 45 U/L in pleural fluid ADA had sensitivity of 72.4% and 76.10% with specificities of 81.53% and 100% respectively, in tuberculous pleural effusion patients. It has been reported that with an ADA activity cut off value of 54 U/L, the sensitivity is 82% and specificity is 97% for the diagnosis of tuberculosis ⁽¹⁵⁾. (Baganha et al, 1990).

Reported cut off value for ADA (total) vary from 47 to 60 U/L (valeses et al 1993⁽¹⁶⁾, perez-Rodriguez et al, 1999⁽¹⁷⁾, Roth et al, 1999⁽¹⁸⁾).

Several studies have suggested that an elevated pleural fluid ADA level predict tuberculous pleuritis with a sensitivity of 90-100% and specificity of 89-100% where the guisti method (colorimetric) is used ⁽¹⁹⁾.

Conclusion

Pleural and peritoneal effusions occur in a large variety of pathological condition. Different laboratory tests on fluid have been advocated but they all have shown either less sensitivity or time consuming.

This study shows very encouraging results with 98.5% sensitivity & 98.57% specificity of ADA test for the diagnosis of tuberculosis infection from pleural, peritoneal & Cerebrospinal fluid.

Many sensitive tests involving molecular diagnosis is available for rapid diagnosis of TB, but they are expensive, require advanced infrastructure and expertise skill, which is not available in developing countries like India. The colorimetric method for the measurement of total adenosine deaminase (ADA) activity has advantages over other methods because of its lower cost, simplicity of technique and rapid turn- around time. The estimation of ADA activity in body fluids such as pleural, peritoneal & CSF there for serves as a reasonable tool in the diagnosis of Tuberculous pleural and peritoneal effusion & TB meningitis when other clinical laboratory tests are negative.

Reference

1. Dye.c., Scheele, s., pathania v and Raviglione, M.C. Global burden of tuberculosis: estimated incidence, prevalence and mortality by country. JAMA 282, 677, 1999.
2. World Health Organization. Global tuberculosis control, surveillance, planning , financing. WHO /CDS/ Tuberculosis/2002, 295.
3. World Health Organization. Prevalence and incidence of tuberculosis in India: A Comprehensive Review, 1997, WHO/TB/97, 231, 1998.
4. Progress towards tuberculosis control India 2001.MMWR 51:229, 2002.
5. Global TB/HIV working group. Tuberculosis and HIV. [http:// www.who.int/en](http://www.who.int/en)
6. Light RW pleural diseases.4th ed. Philadelphia, Lippincott William & Wilkin: 2001.
7. Gupta VK, Mukerji s, Datta SK. Diagnostic evvaluation of ADA in tuberculosis peritonitis.JAPI 1992; 40(6) :387-389.
8. Voight MD, Kalvaria L, Trey C, *et al.* Diagnostic value of ascites adenosine deamainase activity in tuberculous peritonitis.Lancet- 1989; 1:751-754.
9. Dwivedi M, Misra SP, Misra V,at el. Value of adenosine deaminase estimation in the diagnosis of tuberculous ascites. AM J Gastroenterology 1990;85:1123-5.
10. Text book of biochemistry with clinical corelation. Devline Thomas M 6th edition
11. Sharma SK, suresh V, Mohan A,et al. A prospective study of sensitivity and specificity of adenosine deaminase in the diagnosis of tuberculosis pleural effusion. Indian j chest Dis Allied Sci 2001;43:149-55.
12. Gupta DK. Suri JC, Goel A. Effecasy of ADA in diagnosis of pleural effusion.Ind j chest Dis 1990;32(4):205-208.
13. Burgess L.J.use of adenosine deamianase as a diagnostic tool for tuberculous pleurisy. Thorax 1995 june ;50(6); 672-674.
14. Shibagaki T.et al. Adenosine deaminase isozymes in tuberculous pleural effusion.J Lab clin Med 1996 April,127(4):348-352.
15. Baganha MF, pego A, Lima MA, Gaspar EV, Cordeiro AR. Serum and pleural adenosine deamanase : correlation with lymphocytes population. Chest 1990; 97:605-10.
16. Valdes L, san jose E, Alvarez D,et al. Diagnosis of tuberculous pleurisy using the biologic parameters adenosine deamanase, Lysozyme and Interferon gamma. Chest 1993;103:458-65.

17. Perez-Rodriguez E, Perez Walton IJ, Sanchez Hernandez JJ, et al. ADA₁/ADA P ratio in pleural tuberculosis : an excellent diagnostic parameter in pleural fluid, *Respir Med* 1999;93:816-21.
18. Roth B J. searching for tuberculosis in the pleural space. *Chest* 1999;116:3-5.
19. Guistii G, Galanti B. Colorimetric method. In: Bergmeyer HV, ed. *Methods of enzymatic analysis*. Weinheim: Verlag Chemie, 1984:315-23.