



ISSN 2278 – 0211 (Online)

## Prevalence of *Salmonella* in Meat of Food Animals

**Kiran**

M. V. Sc. Student, College of Veterinary & Animal Sciences  
G. B. Pant University of Agriculture & Technology, Pantnagar, Uttarakhand, India

**A. K. Upadhyay**

Professor, VPE, College of Veterinary & Animal Sciences  
G. B. Pant University of Agriculture & Technology, Pantnagar, Uttarakhand, India

**Maansi**

Assistant Professor, College of Veterinary & Animal Sciences  
G. B. Pant University of Agriculture & Technology, Pantnagar, Uttarakhand, India

**Anil Kumar**

Veterinary Officer, College of Veterinary & Animal Sciences  
G. B. Pant University of Agriculture & Technology, Pantnagar, Uttarakhand, India

**Abstract:**

An attempt was made to isolate and identify *Salmonella* bacteria from meat samples of different animals around Pantnagar area. A total of 716 meat samples were screened by conventional cultural method and further confirmed by biochemical, serological and PCR based molecular technique. The samples were plated on various media like HEA, BGA, XLD, BSA and MLA from where the putative isolates were tested biochemically followed by serological identification by latex agglutination kit and molecular confirmation using specific genes. A total of 9 salmonella could be isolated.

**1. Introduction**

*Salmonella* is the second major cause of food borne diseases in the world. Over one million and twenty-seven thousand cases of food borne illness are caused by nontyphoidal *Salmonella*, which resulted in 19,336 hospitalizations and 378 deaths in USA alone. Currently, there are more than 2579 serotypes of *Salmonella enterica*. In India, the most frequently associated serotype among non typhoidal *Salmonella* in chicken meat is *S. Typhimurium* (87.8%). Information on prevalence of *Salmonella* in India is fragmentary and no systematic literature is available on prevalence of *Salmonella* from production to consumption level which is essential in understanding the dynamics of *Salmonella* contamination. Therefore, the present investigation was carried out to study the prevalence of *Salmonella* in meat of different animal species and human stool.

**2. Material Method**

During the period of September 2014- February 2015, a total of 716 meat samples comprising of poultry (289), pork (127), chevon (162), fish (97) and carabeef (41) were collected from the district Nainital and Udham Singh Nagar of Uttarakhand state of India and processed on medium as Brilliant Green Agar (BGA), Xylose Lysine Deoxycholate (XLD) Hekoen Enteric Agar (HEA) plates and Bismuth Sulphite Agar (BSA). Morphological (Old, 1996), biochemical (Chandrashekhar, 2012) and serological (Kumar, 2013) characterization of *Salmonella* isolates was carried out.

*Salmonella* isolates identified on the basis of cultural, morphological, biochemical reactions and agglutination tests were sent for serotyping to *Salmonella* Typing Centre, Division of Bacteriology and Mycology, IVRI, Izatnagar, Bareilly (U.P.) Further, isolation of DNA from pure culture was undertaken using Hi-pura DNA extraction kit (Hi-media), following the manufacturer's instructions supplied along with kit. These isolates were then subjected to PCR based molecular characterization. All the suspected isolates got confirmed with PCR and yielded products at 244 bp, 617 bp and 220 bp for the primer pairs targeting *invA*, *stn* and *sopB* gene respectively. The amplified PCR products were analyzed using horizontal submarine gel electrophoresis in 1.8% agarose gel.

**3. Results and Discussion**

In this study, a total of 9 isolates of *Salmonella* were obtained from 716 meat samples of different animal species and 50 human stool samples. In meat, the prevalence rate of *Salmonella* in Pantnagar and nearby areas based on the above findings was calculated as 1.17%, but lower prevalence of 0.94% was reported by Chandrashekhar, (2012). The prevalence rate of *Salmonella* in chicken meat

was recorded as 2.42% concomitant with Kumar and Lakhera, (2013) who reported 2% of chicken meat samples positive for *Salmonella*. While in pork it was calculated to be 1.57%, although in another Danish study, which estimated prevalence on pig carcasses between 0.9–1.2percent (Nielsen *et al.*, 2001).

All the chevon (162) samples failed to reveal the presence of *Salmonella* during the course of the study agreeing with a study on 600 samples examined from goats, only 4 (0.7%) were *Salmonella* positive (Molla, 2006). None of the fish samples were found to be positive for *Salmonella* in the present study, however, in India Hatha and Lakshmanaperumalsamy (1997) reported 14.25% of the fish samples and 17.39% of crustacean samples contaminated with *Salmonella*. The absence of *Salmonella* in fish might be due to the different epidemiological conditions prevailing in this area. The prevalence of *Salmonella* in carabeef was found to be 0% which is in agreement of the study conducted in Canada, where none of the carabeef samples (n= 134) tested were positive for *Salmonella* (Aslam *et al.*, 2012). There was a significant finding that *Salmonella* organism could not be recovered from stool samples of human in Pantnagar. In a study of 232 samples examined, only one sample (0.43%) yielded *Salmonella* serovar (Bisht, 2010).

The overall prevalence of *Salmonella* in different meat samples was 1.17%. Higher prevalence was reported in chicken meat (2.42%) followed by pork (1.57%), as these are the main reservoirs of *Salmonella* bacteria. Both poultry and porcine meats are generally considered higher risks than other meats for *Salmonella* (Duffy *et al.*, 1999). The reason of such low prevalence could be the use of antibiotics and other feed additives given regularly to the animals. Further, samples collected from healthy individuals, less number of processed samples, low endemicity of salmonellosis in this area and good manage mental and hygienic practices might be the other reasons for these observations.

S.No.	Source of sample	No. of processed samples	Total no. of isolates recovered	Serovars identified
1.	Chicken meat	289	7 (2.42%)	<i>S. Typhimurium</i> (6) <i>S. Enteritidis</i> (1)
2.	Pork	127	2 (1.57%)	<i>S. Weltevreden</i> (2)
3.	Chevon	162	-	-
4.	Fish	97	-	-
5.	Carabeef	41	-	-

Table 1: Prevalence of *Salmonella* in different samples

In this study, a total of 9 isolates of *Salmonella* were recovered with maximum prevalence were *S. Typhimurium* (66.66%) followed *S. Weltevreden* (22.22%) and *S. Enteritidis* (11.11%) as depicted in Table: 2 in agreement with data in the European Union, in which *S. Typhimurium* and *S. Enteritidis* was reported as the most prevalent serotype in broiler carcasses in 2009 (EFSA, 2011). These two serotypes are commonly reported from poultry in the Indian subcontinent (Murugkar *et al.* (2005). Since *S. Typhimurium* can infect man and animals equally, interspecies sharing of this serovar have also been observed (Singh *et al.*, 2010). The dominant serotypes identified from beef, poultry and pork meat were *S. Typhimurium* (12.7%), followed by *S. Enteritidis* (12.5%) (Roseliza, *et al.*2011).

S. No	Source	Serovars Identified	Antigenic Structure
1.	Poultry	<i>S. Typhimurium</i>	4,5,12:i:1,2
2.	Poultry	<i>S. Typhimurium</i>	4,5,12:i:1,2
3.	Poultry	<i>S. Typhimurium</i>	4,5,12:i:1,2
4.	Pig	<i>S. Weltevreden</i>	9,12:-:-
5.	Poultry	<i>S. Enteritidis</i>	9,12:gm:-
6.	Poultry	<i>S. Typhimurium</i>	4,5,12:i:1,2
7.	Poultry	<i>S. Typhimurium</i>	9,5,12:g,m:-
8.	Pig	<i>S. Weltevreden</i>	9,12:g,m:-
9.	Poultry	<i>S. Typhimurium</i>	4,5,12:i:1,2

Table 2: *Salmonella* serovars recovered from the samples.

#### 4. Summery

Out of 716 samples comprising of chicken meat (289), pork (157), chevon (162), fish (97) and carabeef (41) collected from the areas of Nainital and Udham Singh Nagar, only 9 could divulge *Salmonella* spp. by conventional cultural method and confirmed by biochemical, serological and PCR based molecular techniques. Six *S. Typhimurium*, two *S. Weltevreden* and only one *S. Enteritidis* could be isolated.

#### 5. Acknowledgement

We are thankful to ICAR for grant of Outreach programme on zoonotic diseases, in which the research was carried out.

**6. References**

- i. Aslam, M., Checkley, S., Avery, B., Chalmers, G. and Bohaychuk, V. 2012. Phenotypic and genetic characterization of antimicrobial resistance in Salmonella serovars isolated from retail meats in Alberta, Canada. *Food Microbiol.* 32 : 110-117.
- ii. Bisht, G. 2010. Isolation, identification and molecular characterization of Salmonella serovars recovered from man and animals. M.V.Sc thesis submitted to Department of Vet. Public Health. Govind Ballabh Pant University of Agriculture & Technology.
- iii. Chandrashekhar, 2012. Isolation, identification and molecular characterization of Salmonellae from man, animals and foods of animal origin with special reference to determination of virulence genes. P.H.D thesis submitted to Department of Vet. Public Health. Govind Ballabh Pant University of Agriculture & Technology.
- iv. Duffy, G., Cloak, O.M., O'Sullivan, M.G., Guillet, A., Sheridan, J.J., Blair, I.S. and McDowell, D.A. 1999. The incidence and antibiotic resistance profiles of Salmonella spp. on Irish retail meat products. *Food Microbiol.* 16: 623– 631.
- v. EFSA. 2011. The European Union summary report on trends and sources of zoonoses.
- vi. Hatha, A.A.M. and Lakshmanaperumalsamy, P. 1997. Prevalence of Salmonella in fish and crustaceans from markets in Coimbatore, South India. *Food Microbiol.* 14: 111–116.
- vii. Kumar, K and Lakhera, P.C. 2013. Isolation, plasmid profiling and antibiogram of Salmonella from poultry meat and env. sources. *J. of Anim. Res.* 3: 53-57.
- viii. Molla, W., Moll, B., Alemayehu, D., Muckle, A., Cole, L. and Wilkie, E. 2006. Occurrence and antimicrobial resistance of Salmonella serovars in apparently healthy slaughtered sheep and goats of central Ethiopia. *Trop. Anim. Health Prod.* 38: 455–462.
- ix. Murugkar, H.V., Rahman, H., Kumar, A. and Bhattacharyya, D. 2005. Isolation, Phage typing and antibiogram of Salmonella from man and animals in northeastern India. *Ind. J. Med. Res.* 122: 237-242.
- x. Nielsen, B., Alban, L., Stege, H., Sørensen, L.L., Møgelmoose, V., Bagger, J., Dahl, J. and Baggesen, D.L. 2001. A new Salmonella surveillance and control programme in *Wochenschrift.* 114: 323–326.
- xi. Old, D.C. 1996. Salmonella. In: Mackie and McCartney *Practical Medical Microbiology.* 14<sup>th</sup> Edn. Churchill Livingstone Edinburgh London New York Philadelphia Sydney Toronto.
- xii. Roseliza, R., Maswati, M.A., Hasna, Y. and Ramlan, M. 2011. Identification of Salmonella serotypes isolated from meat samples in Malaysia. *Malaysian J. of Vet. Res.* 2: 59-64.
- xiii. Singh, S, Agarwal, R.K., Tiwari, S.C., Kumar, K. and Nambiar, K. 2010. Predominance of Paratyphi B var Java serotype among human and animals Clin. isolates of Salmonella in North India. *J. Vet. Pub. Hlth.* 8(2): 73-81.